

Bacteriocins and Bacteriophage Lytic Proteins as Alternatives to Antibiotics from Russian Federation and USA Collaborations

Bruce S. Seal¹, Nikolay V. Volozhantsev², J. Eric Line¹, Edward A. Svetoch¹, Gregory R. Siragusa^{1*}, Norman J. Stern¹

¹Poultry Microbiological Safety Research Unit, Richard B. Russell Agricultural Research Center, ARS, USDA, Athens, GA 30605 USA

²State Research Center for Applied Microbiology & Biotechnology, Obolensk, Moscow Region, Russian Federation



Session 1

ABSTRACT

Novel anti-microbial peptides (bacteriocins) were isolated and characterized during collaborative research between PMSRU, ARS-USDA scientists and representatives of the State Research Center for Applied Microbiology and Biotechnology (SRCAMB) in Obolensk, Russian Federation. The bacteriocins are effective against several bacterial pathogens. Treatment of chickens by feeding bacteriocins consistently reduced Campylobacter levels in their gastrointestinal system as compared with levels found in untreated birds. Five patents have been issued describing this alternative to antibiotics treatment for bacterial infection and technology transfer is ongoing. Screening of bacteriophages lytic for Clostridium perfringens was completed utilizing filtered samples obtained from poultry (intestinal material), soil, sewage and poultry processing drainage water. From the collections highly lytic viruses were isolated and the double-stranded deoxyribonucleic acid (DNA) genomes of the bacteriophages were sequenced to completion. DNA sequencing of six bacteriophage genomes completed at PMSRU and four genomes in collaboration with Russian investigators resulted in identification of unique amidases as well as phage encoded proteins that potentially contain lysozyme and endopeptidase activities. Two recombinant bacteriophage lytic enzyme genes encoding putative amidases have been cloned, their proteins expressed as recombinants and isolated to homogeneity, then demonstrated to lyse *C. perfringens*. Patent applications have been submitted as a result of the bacteriophage research. These bacteriocins and phage lytic enzymes may have possibilities for use in agriculture and medical applications as potential replacements for current antibiotics that may have diminished activity.

INTRODUCTION

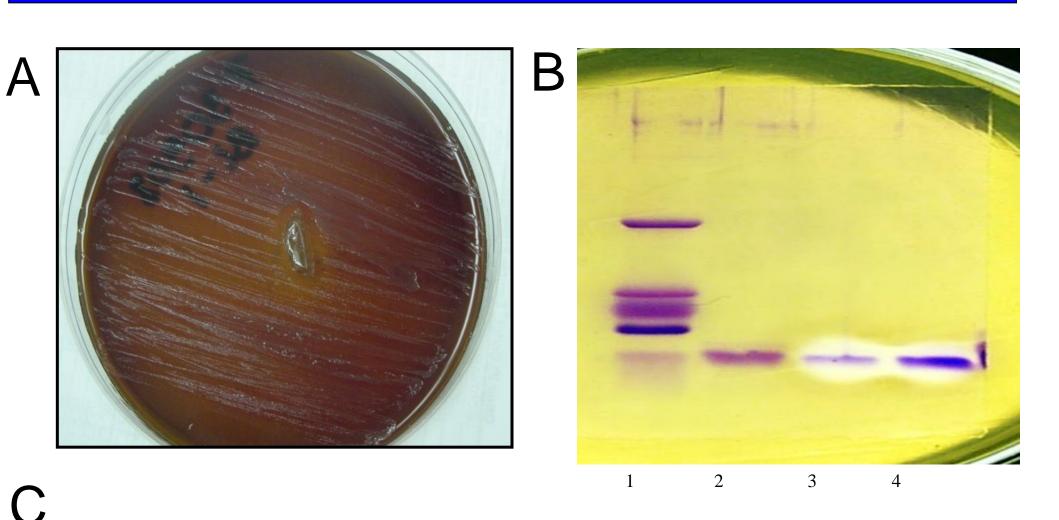
The World Health Organization (WHO) reports that governments worldwide are intensifying their efforts (http://www.who.int/mediacentre/factsheets/fs237/en/) to improve food safety. Internationally food-borne disease is difficult to estimate, but it has been reported that in 2005 alone 1.8 million people died from diarrheal diseases and a great proportion of these cases were attributed to contamination of food and drinking water (WHO, 2010 web site). The majority of pathogens causing this significant disease burden are considered to be zoonotic (Schlundt et al., 2004). The most important factors driving an increase in the burden of food-borne disease during the future are anticipated to be a doubling of the global demand for food accompanied by increased international trade in food. This will be accompanied by an increase in consumption of certain high-value food commodities such as meat, poultry and fresh produce. One of the most important factors in reducing the burden of food-borne disease was identified as development of effective control measures (Quested et al., 2010). The USA Centers for Disease Control and Prevention estimates that in the United States alone Campylobacter spp., Salmonella spp. (non-typhoid), and CPE-producing Type A Clostridium perfringens, are the three leading bacterial etiologies of human food-borne illness with 1.32 million, 1.23 million and 0.97 million domestic cases respectively (Scallan et al., 2011). The application of therapeutic bacteriocin treatments to reduce poultry colonization diminishes Campylobacter spp. in cecal material to low levels in treated birds and this could be a valid means for controlling food-borne bacteria in poultry (Svetoch and Stern, 2010). Also, bacteriophage gene products may be another avenue for developing alternative antimicrobials to control pathogenic bacteria (Liu et al., 2004).

METHODS

Bacteriocins were purified from lactic acid bacteria isolated from chicken gastrointestinal materials. The bacteriocins were then assayed for their antimicrobial properties (Line et al., 2008; Stern et al., 2006; Svetoch et al., 2011). Bacteriophages that infected *C. perfringens* were isolated from poultry intestinal material, poultry processing offal and sewage (Oakley et al., 2011; Volozhantsev et al., 2011; Volozhantsev et al., 2011; Volozhantsev et al., 2012). Recombinant lytic proteins were subsequently expressed and assayed for the ability to lyse *C. perfringens* (Simmons et al., 2010).

Email contact information for authors: bruce.seal@ars.usda.gov; svetoch@obolensk.org; nikkvol@oblensk.org; eric.line@ars.usda.gov *Present address: Danisco-DuPont, Waukesha, WI 53188 USA

RESULTS



OR7 1 KTYYGTNGVHCTKNSLWGKVRLKNM-----KYDQNTTYMGRLQDILLGWATGAFGKTFH 54
KTYYGTNGVHCT K SLWGKVRLKN + + + + + ILLGWATGAFGKTFH
acidocin A 24 KTYYGTNGVHCTKKSLWGKVRLKNVIPGTLCRKQSLPIKQDLKILLGWATGAFGKTFH 81

OR7 peptide amino acid composition: 28.4% charged (RKHYCD); 3.3% acidic (DE); 13.3% basic (KR); 31.6% polar (NCQSTY); 23.4% hydrophobic (AILFWV);no glutamine (E) or proline (P)+

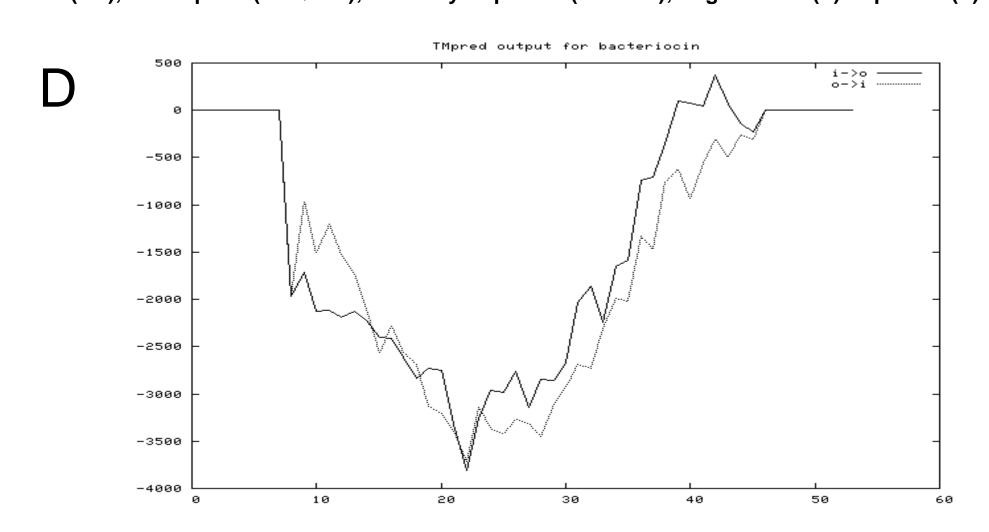


Fig 1. Bacteriocin characteristics (A) Lactic acid bacteria selected for inhibition of *Campylobacter jejuni* growth. (B) SDS-PAGE of purified bacteriocin with bacterial overlay. (C) Alignment and biochemical characteristics of common bacteriocins. (D) Transmembrane domain prediction for bacteriocin OR7.

Table 1. Ten chicks/group were colonized on day of hatch with 10¹⁰ CFU *Campylobacter jejuni* and treated with OR-7 for 10 days.

Trial and Treatment	Campy challenge strain	Treatment days	Mean log ₁₀ CFU/gm cecum
1 Control	AL-22	0	7.2 <u>+</u> 0.3
1 Treated	AL-22	7-9	ND
2 Control	BH-6	0	7.1 <u>+</u> 0.4
2 Treated	BH-6	7-9	0.7 <u>+</u> 1.2
3 Control	BL-1	0	7.8 <u>+</u> 0.2
3 Treated	BL-1	7-9	1.3 <u>+</u> 1.8
4 Control	CL-11	0	6.6 <u>+</u> 0.7
4 Treated	CL-11	7-9	ND

NOTE: During chicken feeding trials, bacteriocin treatment significantly reduced the numbers of *C. jejuni* organisms compared to those found in the untreated control groups of birds (*P* ≤ 0.05; ND-not detected).

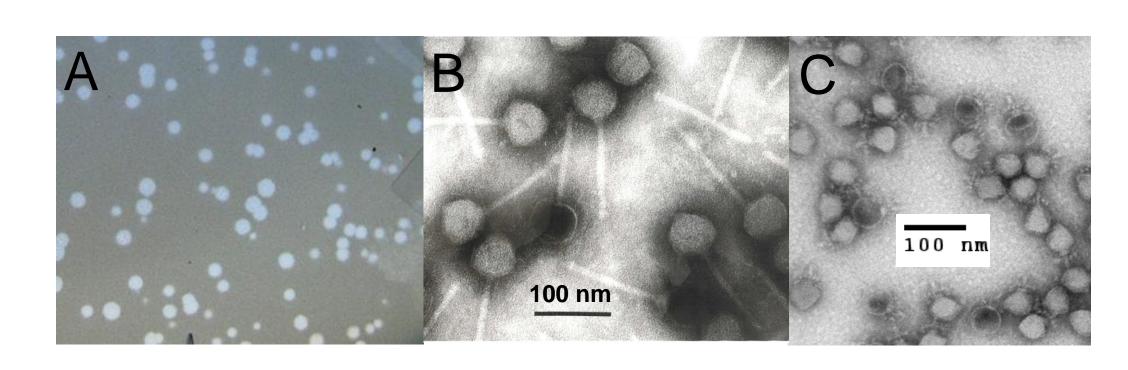


Fig 2. Representative clear plaques and electron micrographs for Russian and USA bacteriophages. (A) All phages were plaque purified at least 3X. (B) Many phages had long non-contractile tails representative of the *Siphoviridae*. (C) Several phages were representative of the *Podoviridae* with short non-contractile tails.

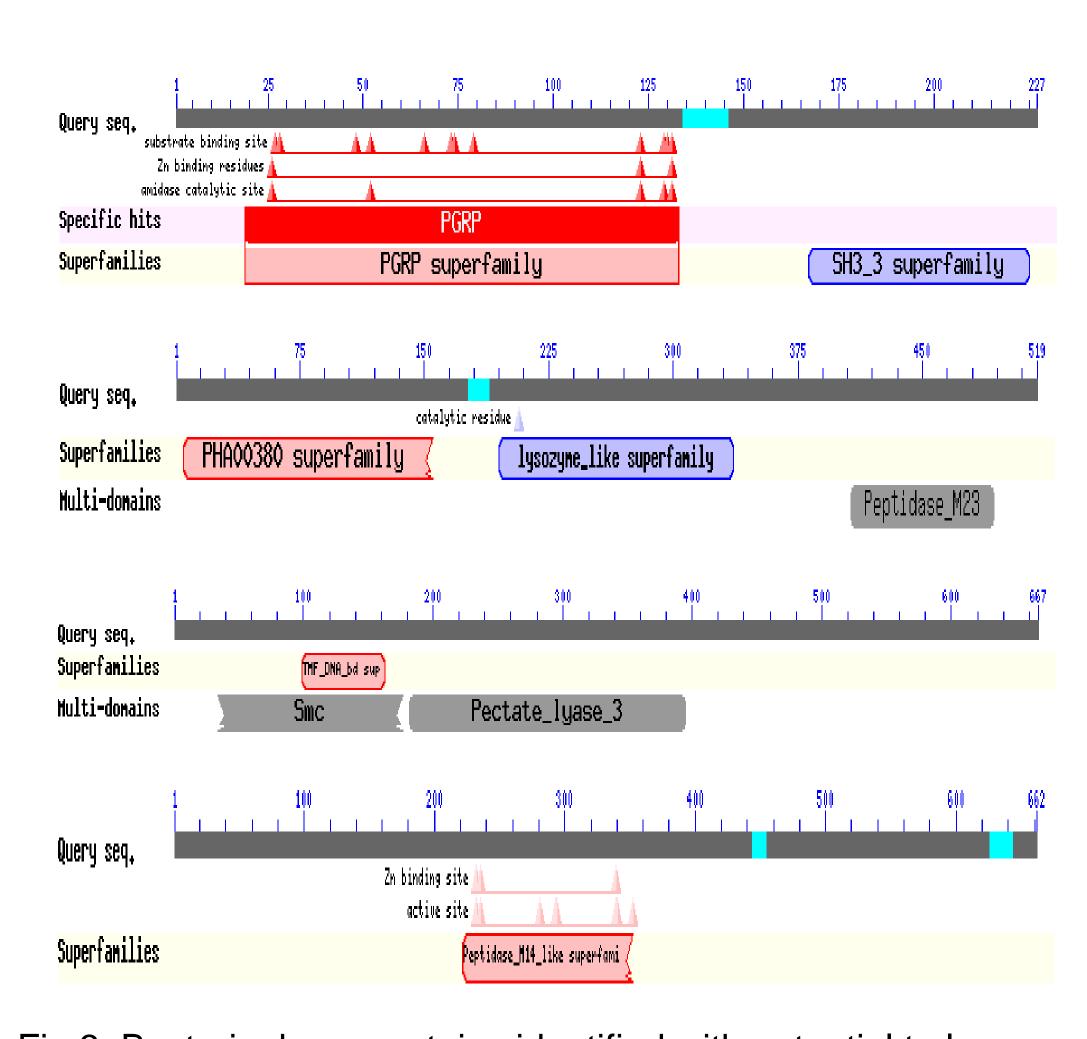
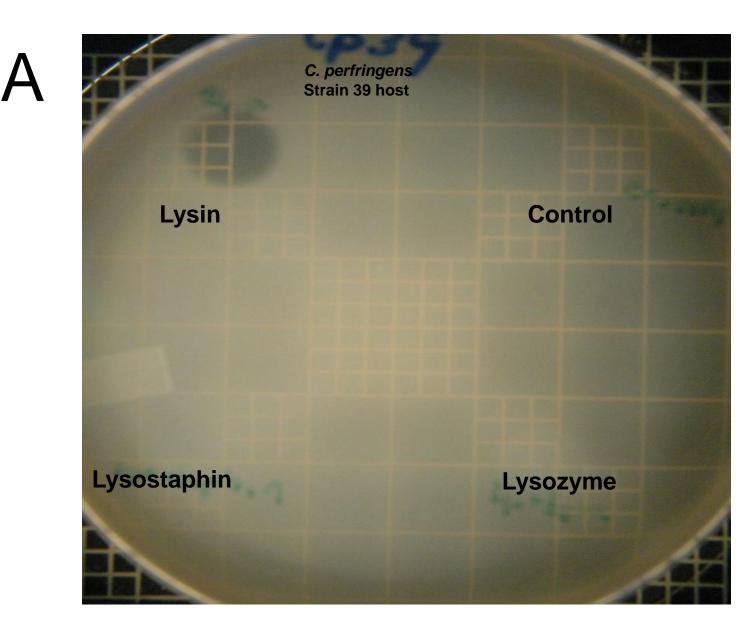


Fig 3. Bacteriophage proteins identified with potential to lyse *Clostridium perfringens*. Includes N-acetylmuramoyl-L-alanine amidases with C-terminal cell wall binding domains, tail proteins with lysozyme and peptidase activities as well as a pectate lyase and a previously unknown viral Zn-endopeptidase.

RESULTS CONTINUED



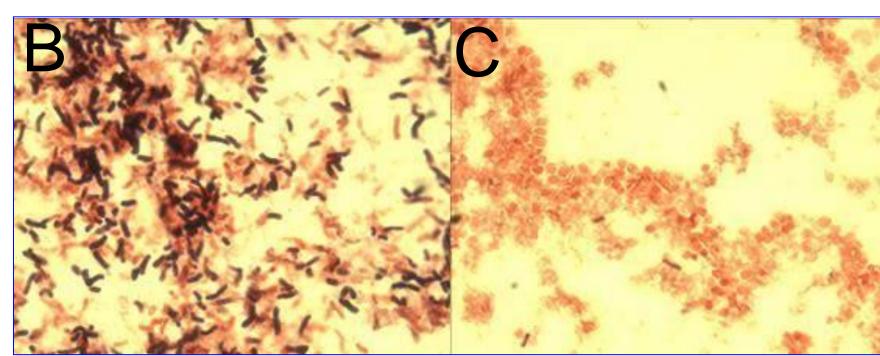


Fig 4. Spot lysis assay of PlyCP39O/CP26Fand Gram stain of *C. perfringens* lysin-treated cells. (A) Spot assay and (B) cells treated without phage lysin or (C) with lysin.

CONCLUSIONS

- Bacteriocins were discovered with activity against the food-borne pathogen *Campylobacter jejuni*.
- Bacteriophages encoded proteins as potential antimicrobials to lyse *Clostridium perfringens*.
- Future need is expression in yeast with large-scale production for field trials in food animals.

REFERENCES

Line JE, Svetoch EA, Eruslanov BV, Perelygin VV, Mitsevich EV, Mitsevich IP, Pokhilenko VD, Levchuk VP, Svetoch OE, Seal BS, Siragusa GR, Stern NJ. 2008. Isolation and purification of enteroicin E-760 with a broad antimicrobial activity against Grampositive and Gram-negative bacteria. Antimicrob. Agents Chemother. 52:1094-1100.

Liu J, Dehbi M, Moeck G, Arhin F, Bauda P, Bergeron D, Callejo M, Ferretti V, Ha N, KwanT, McCarty J, Srikumar R, Williams D, Wu JJ, Gros P, Pelletier J, DuBow M. 2004. Antimicrobial drug discovery through bacteriophage genomics. Nat Biotechnol. 22:185-91.

Oakley BB, Talundzic E, Morales CA, Hiett KL, Siragusa GR, Volozhantsev NV, Seal BS. 2011. Comparative genomics of four closely related *Clostridium perfringens* bacteriophages reveals variable evolution among core genes with therapeutic potential. BMC Genomics. 12:e282.

Quested TE, Cook PE, Gorris LG, Cole MB. 2010 Trends in technology, trade and consumption likely to impact on microbial food safety. Int J Food Micro.139 Suppl1:S29-42.

Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson MA, Roy SL, Jones JL, Griffin PM. 2011. Foodborne illness acquired in the United States--major pathogens. Emerg Infect Dis.17:7-15.

Schlundt J, Toyofuku H, Jansen J, Herbst SA. 2004. Emerging foodborne zoonoses. Rev Sci Tech. 23:513-533.

Simmons M, Donovan DM, Siragusa GR, Seal BS. 2010. Recombinant expression of two bacteriophage proteins that lyse *Clostridium perfringens* and share identical sequences in the C-terminal cell wall binding domain of the molecules but are dissimilar in their N-terminal active domains. J Agric Food Chem. 58:10330-10337.

Stern NJ, Svetoch EA, Eruslanov BV, Perelygin VV, Mitsevich EV, Mitsevich IP, Pokhilenko VD, Levchuk VP, Svetoch OE, Seal BS. 2006. Isolation of a *Lactobacillus salivarius*: its bacteriocin is inhibitory to *Campylobacter jejuni* in chickens. Antimicrob. Agents Chemother. 50:3111-3116.

Svetoch EA, Eruslanov BV, Levchuk VP, Perelygin VV, Mitsevich EV, Mitsevich IP, Stepanshin J, Dyatlov I, Seal BS, Stern NJ. 2011. Isolation of *Lactobacillus salivarius* 1077 (NRRL B-50053) and characterization of its bacteriocin, including the antimicrobial activity spectrum. Appl Environ Microbiol. 77:2749-54.

Svetoch EA, Stern NJ. 2010. Bacteriocins to control *Campylobacter* spp. in poultry--A review. Poult Sci. 89:1763-8.

Volozhantsev NV, Oakley BB, Morales CA, Verevkin VV, Bannov VA, Krasilnikova VM, Popova AV, Zhilenkov EL, Garrish JK, Schegg KM, Woolsey R, Quilici DR, Line JE, Hiett KL, Siragusa GR, Svetoch EA, Seal BS. 2012. Molecular characterization of podoviral bacteriophages virulent for *Clostridium perfringens* and their comparison with members of the *Picovirinae*. PLoS One. 7:e38283.

Volozhantsev NV, Verevkin VV, Bannov VA, Krasilnikova VM, Myakinina VP, Zhilenkov EL, Svetoch EA, Stern NJ, Oakley BB, Seal BS. 2011. The genome sequence and proteome of bacteriophage ΦCPV1 virulent for *Clostridium perfringens*. Virus Res. 155:433-439

ACKNOWLEGEMENTS

These investigations were supported by ARS-USDA CRIS project number 6612-32000-046-0D, the State Research Center for Applied Microbiology & Biotechnology, Obolensk and the U.S. Department of State via the International Science and Technology Center grants #1720, #3108, #3445 administered by the ARS-USDA Office of International Research Programs with the administrative support of Melanie Peterson (ARS, OIRP) and Patrick Russo (ISTC).